

The Chinese University of Hong Kong, Faculty of Medicine,
Department of Microbiology
Joint Graduate Student Seminar

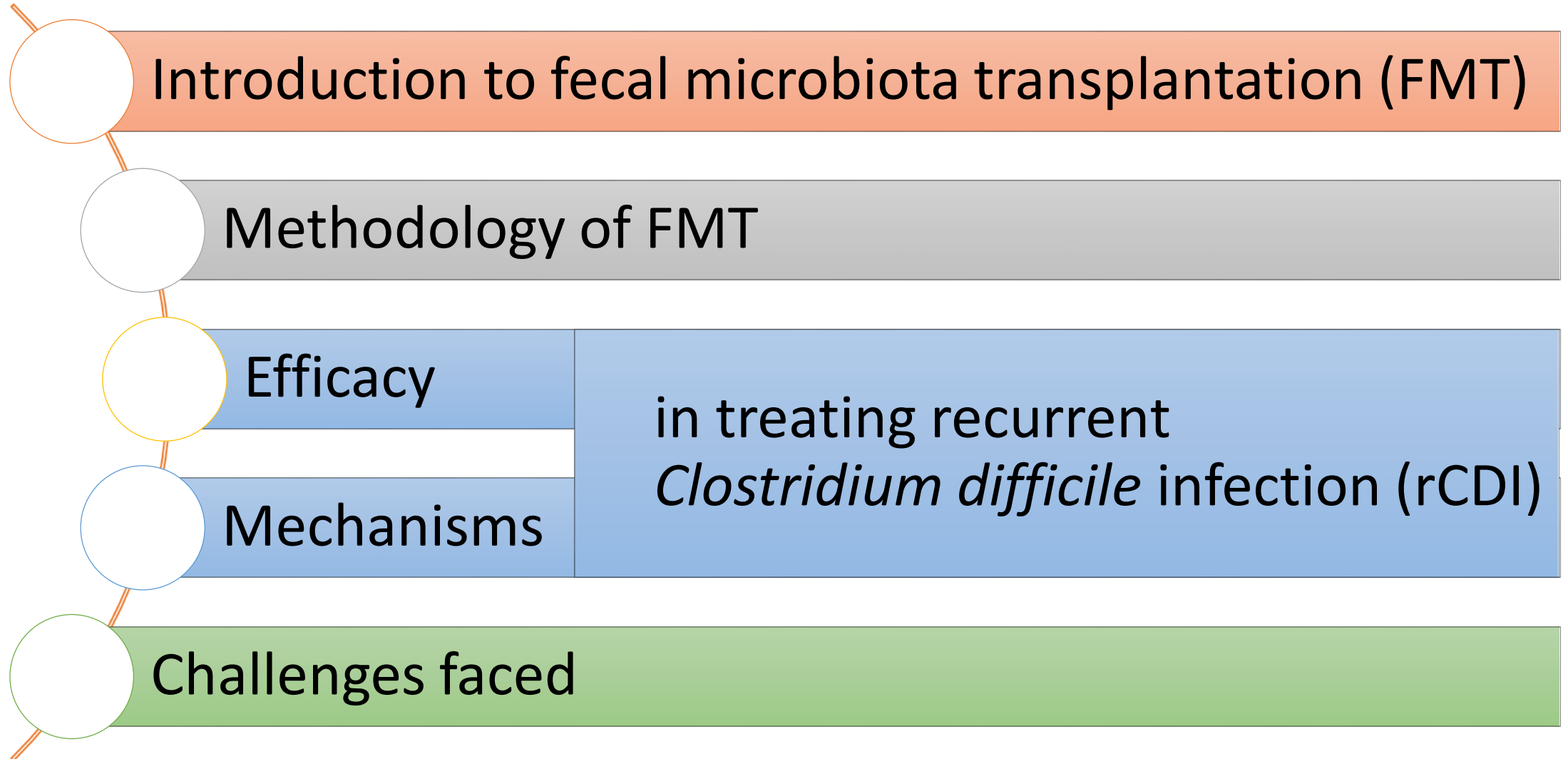
Fecal Microbiota Transplantation: How it's Done, How it Works, and What Challenges Does it Face

Supervisor: Prof. Mamie Hui

Student: Poon Yeuk Lan, Nana (PhD student)

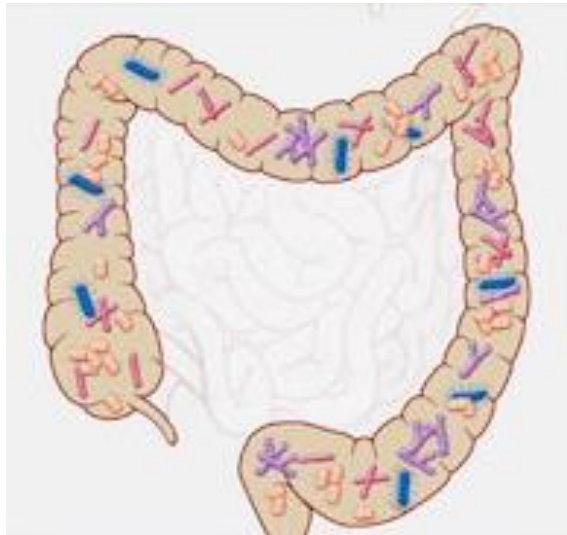
Date: 5th December 2017

Presentation outline



What's FMT?

- Fecal Microbiota Transplantation



Healthy donor

Transplantation of
gut microbiota



Through administration
of fecal material



Patient

To cure the disease or improve the patient's conditions
By normalizing microbial diversity & community structure

History of FMT

1958: First case series of use of FMT

- Eiseman *et al.*, 1958
- Four patients with pseudomembranous enterocolitis (PMC)
- Fecal retention enemas → Prompt recovery in all patients

1983: First documented case of FMT for rCDI

- Schwan *et al.*, 1983
- 65-year-old woman with 5 episodes of CDI relapses
- Fecal enemas
→ Prompt and complete normalization of bowel function

History of FMT

- Since 2000: Booming of FMT practices

Driven by:

1. Increasing CDI incidence

- 6-25% of patients experienced recurrent CDI
- 60% of rCDI patients had multiply recurrent CDI (mrCDI)
- High rate of remission was achieved by FMT

(Cohen et al., 2010)

2. Better understanding in gut microbiota

- Provided logical reasons for using FMT
- Growing acceptance
- Association to other diseases
- FMT applications other than rCDI

Methodology of FMT

Donor
selection

Donor
screening

Stool
preparation

Administration

Methodology of FMT—Donor Selection

Known donors

Patients' spouse, family members or friends

Pros: Better acceptance

Cons: Shared genetic / environmental risk factors
May conceal relevant infectious risk factors

Unrelated, rigorously screened healthy volunteers

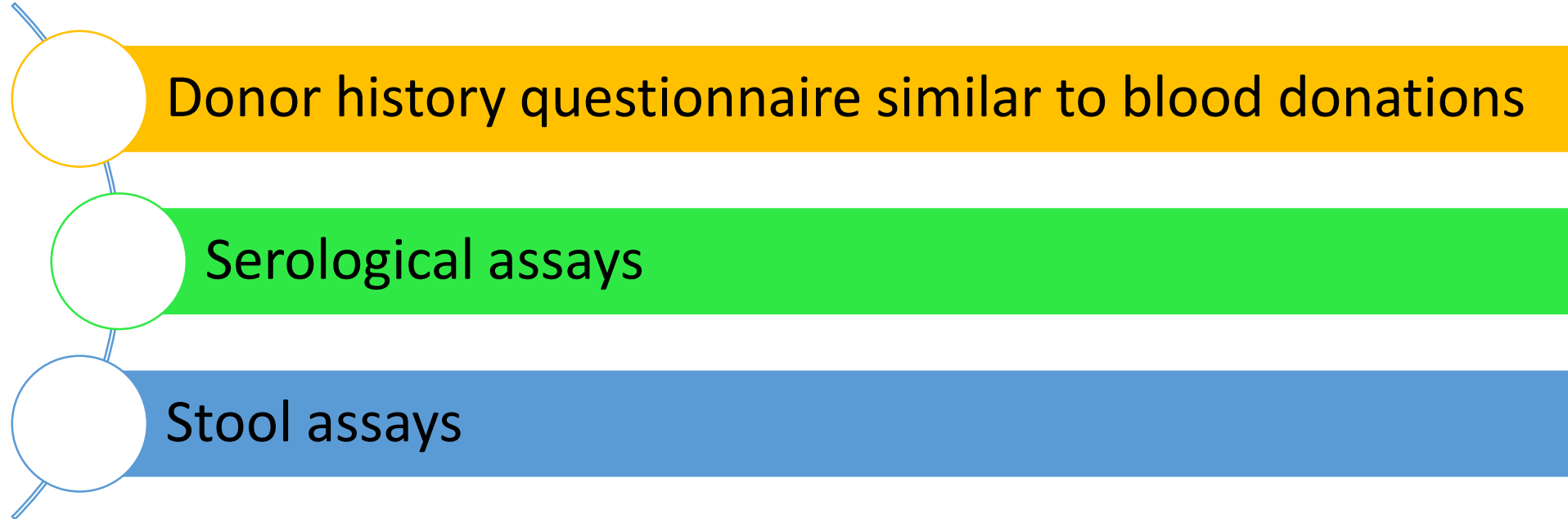
Donations are processed and stored by stool banks

Pros: Do not share genetic / environmental risk factors
Constant ready-to-use supply

Cons: Stool bank may not be available in the patients' region

Methodology of FMT—Donor Screening

- General screening procedures:

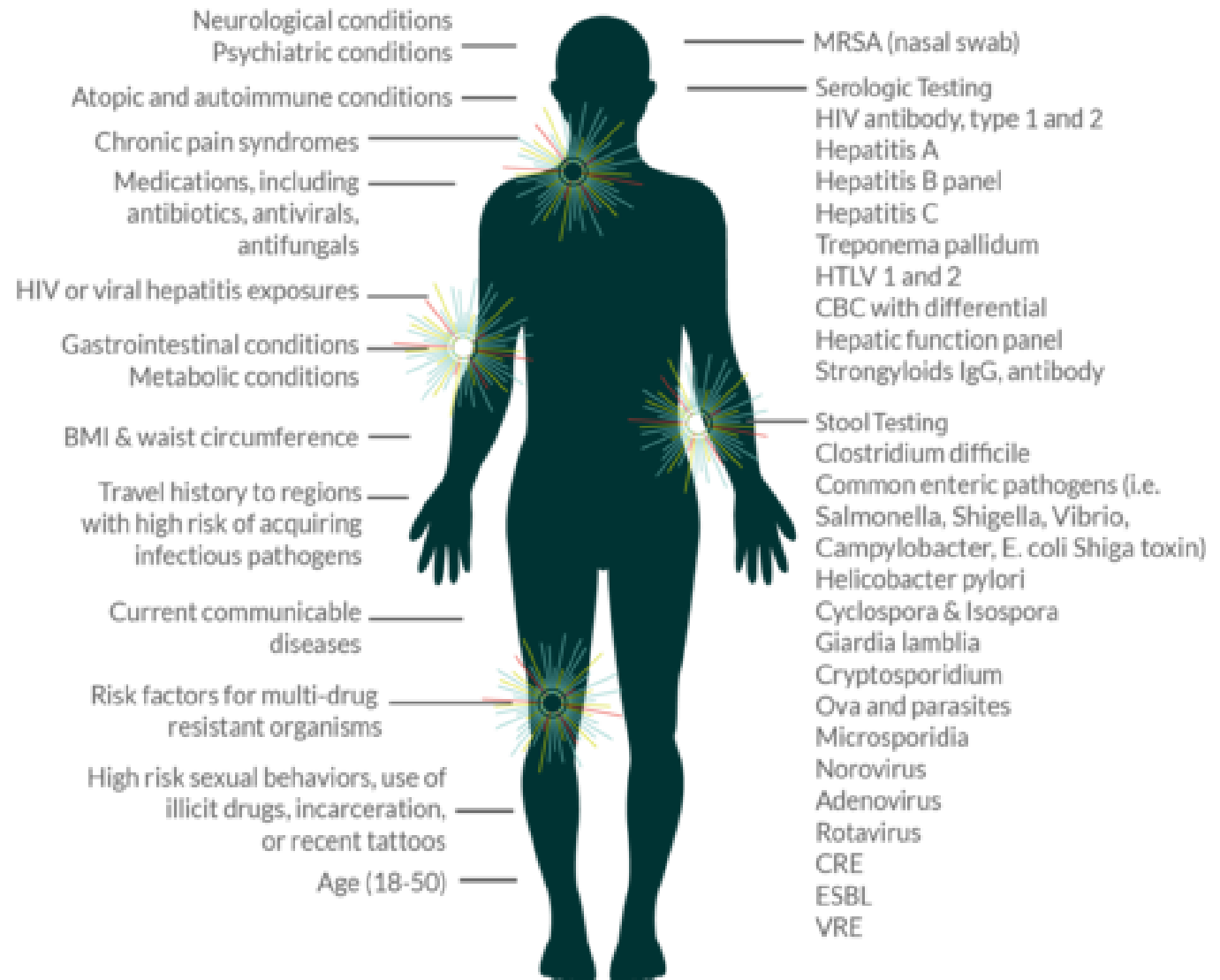


- Screen for diseases or disorders
- Screen for risks of infection

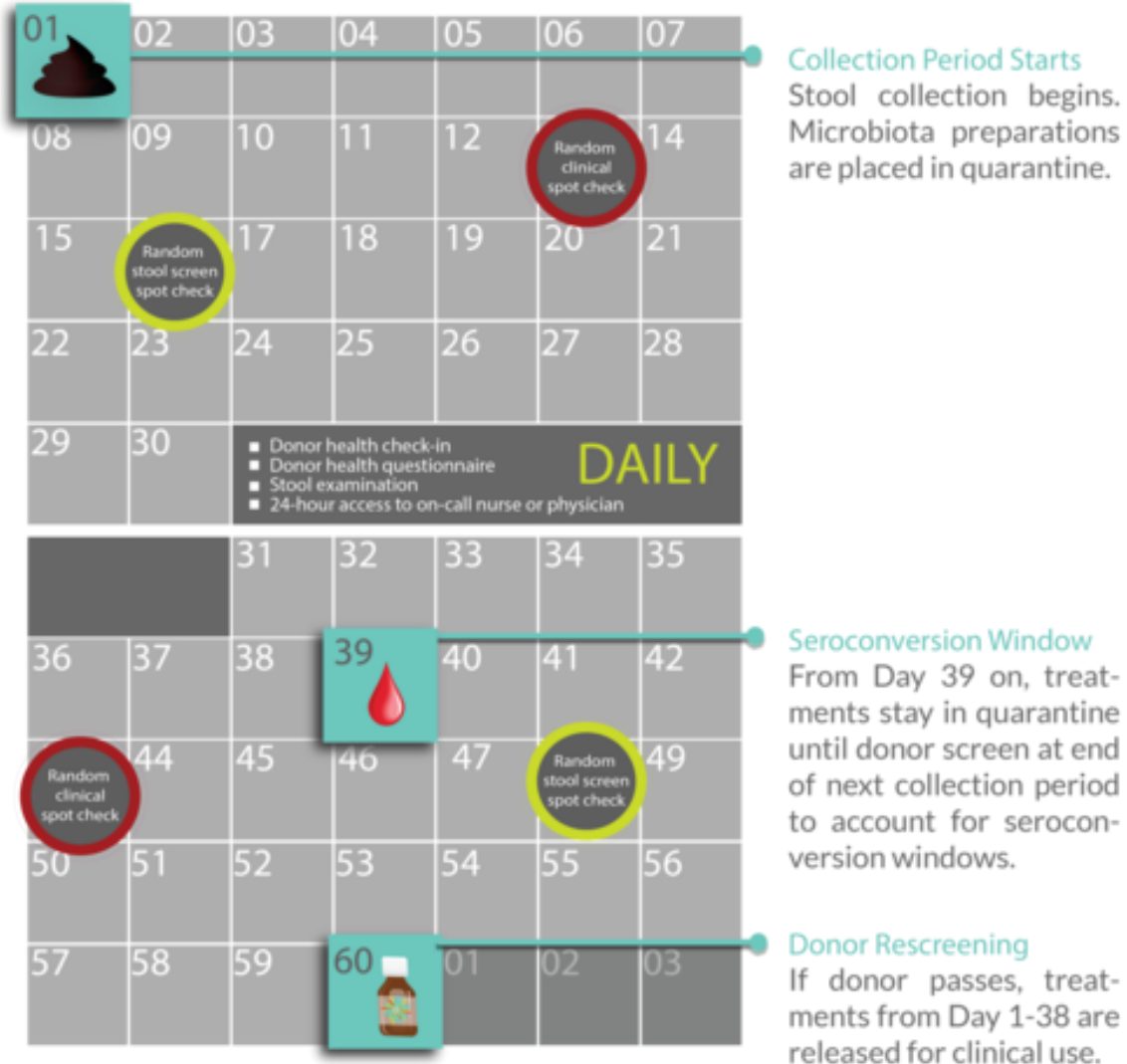
Example of OpenBiome

- OpenBiome:
First public stool bank in USA
- Donor Assessment:
 1. 200-question Clinical Evaluation with internal medicine specialist
 2. Serological assays
 3. Stool-based assays
- Only 3% of volunteers pass and become active donors

Donors are screened for infectious risk factors & potential microbiome-mediated conditions



Example of OpenBiome



- Donors donate stool 3 times a week for ≥ 60 days
- Their health conditions are monitored daily
- After the collection period, they are **rescreened**
- If passed, Stool from Day 1-38 can be released for clinical use
- Stool from Day 39 and on will be released after **the 3rd screening**

Methodology of FMT—Stool preparation

- Stool Collection
 - Stool is collected in sterile containers
 - Delivered for processing within hours



Fisherbrand™ Commode Specimen Collection System

An example of stool collection kit

Methodology of FMT—Stool preparation

- 30~50 g of fecal material are processed
- Homogenization
 - Manual effort
 - Blender
- Diluents
 - Tap water
 - Milk
 - Saline
 - Saline with glycerol



Household blender



Paddle blender

Stomacher® 400
Circulator Lab Blender

(Kelly et al., 2015)

Methodology of FMT—Stool preparation

- Filtration
 - To remove particles
 - To prevent clogging in infusion tube
 - Can be done with gauze, coffee filter or strainer
- Processed sample can be
 - Directly administered to patients
 - Frozen for later use



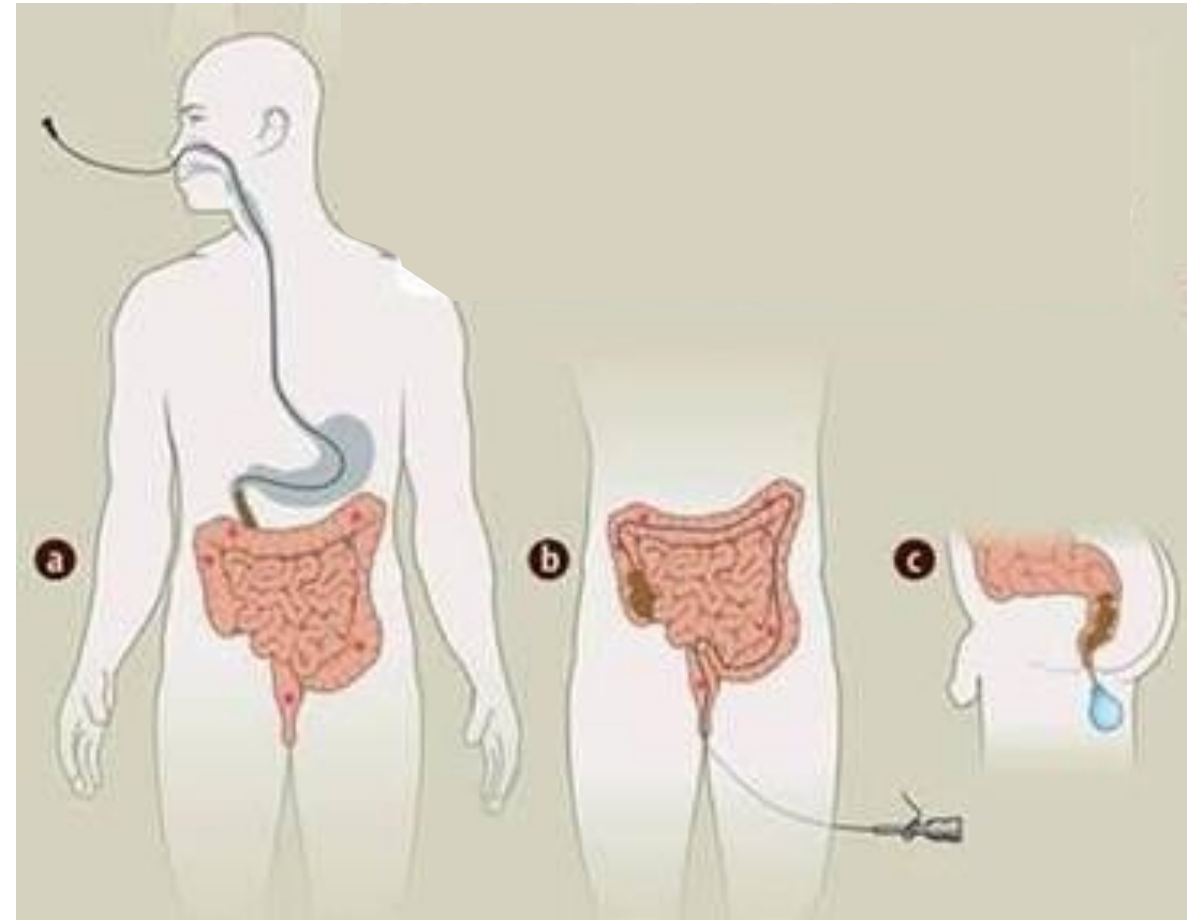
(Kelly et al., 2015)

Methodology of FMT—Administration

- Patient preparation
 - Discontinue antibiotics 12-48 hours before FMT
 - Administer proton pump inhibitor before FMT for upper GI tract and oral delivery

Methodology of FMT—Administration

- Routes of administration
 - Upper GI tract
 - Endoscopy, nasogastric/ nasointestinal tubes
 - 25-50 ml of stool suspension
 - Lower GI tract
 - Colonoscopy, sigmoidoscopy, rectal tube, enema
 - 200-500 ml of suspension



Nasointestinal
tube

Colonoscopy

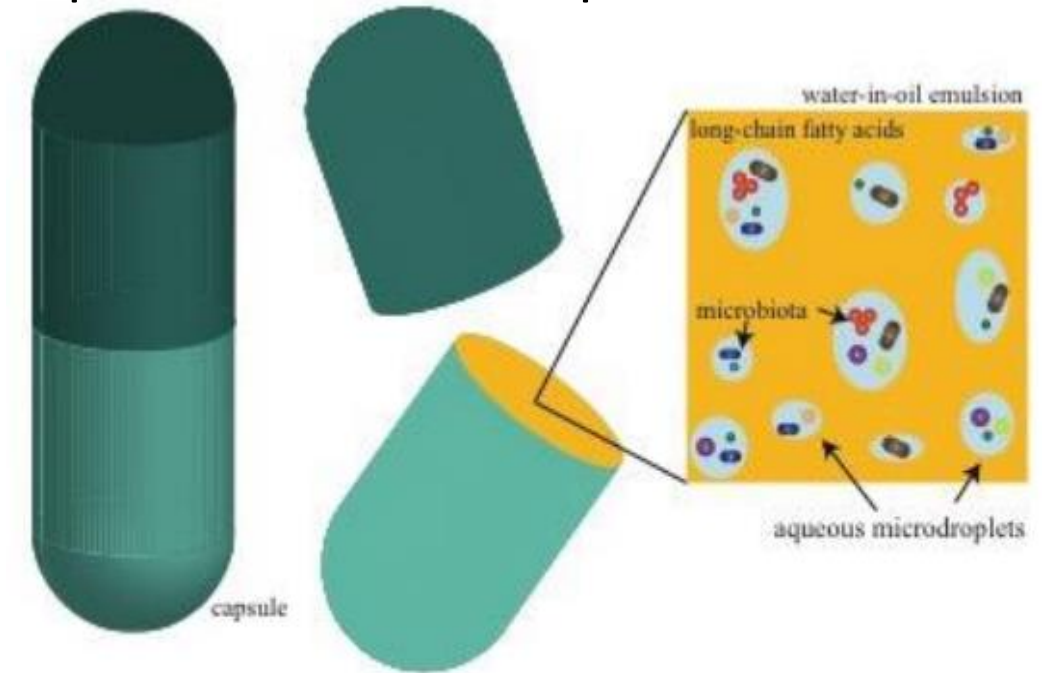
Enema

(Cammarota et al., 2017; Kelly et al., 2015)

Methodology of FMT—Administration

- Routes of administration
 - Oral—Capsules
 - Dose: ~30 capsules
 - Contain same amount of stool material in traditional FMT
 - Advantages:
 - Non-invasive
 - Eliminate procedure-related cost
 - More aesthetically appealing

OpenBiome's FMT Capsule



FMT Capsule G3 Ingredients: Frozen human fecal microbiota filtered to 330 microns, theobroma oil, glycerol, hide bovine gelatin, sodium lauryl sulfate, colorants FD&C, titanium dioxide

Efficacy of FMT in rCDI

- Rapid resolution of symptoms within days
- High cure rate

Systematic review by Li et al. in 2016

- Included 18 cohort and case series studies
- Involved 611 patients
- Long-term follow-up duration (≥ 90 days)
- Pooled primary cure rate: 91.2% (95% CI 86.7–94.8%)
- Overall recurrence rate: 5.5% (95% CI 2.2–10.3%)

Efficacy of FMT in rCDI – Clinical trials

- Two clinical trials comparing efficacy of FMT with vancomycin

van Nood et al., 2013

Resolution of rCDI occurred in

- FMT group: 13/16 (81%) of patients
- Vancomycin group: 4/13 (31%) of patients

Cammarota et al., 2015

Resolution of rCDI occurred in

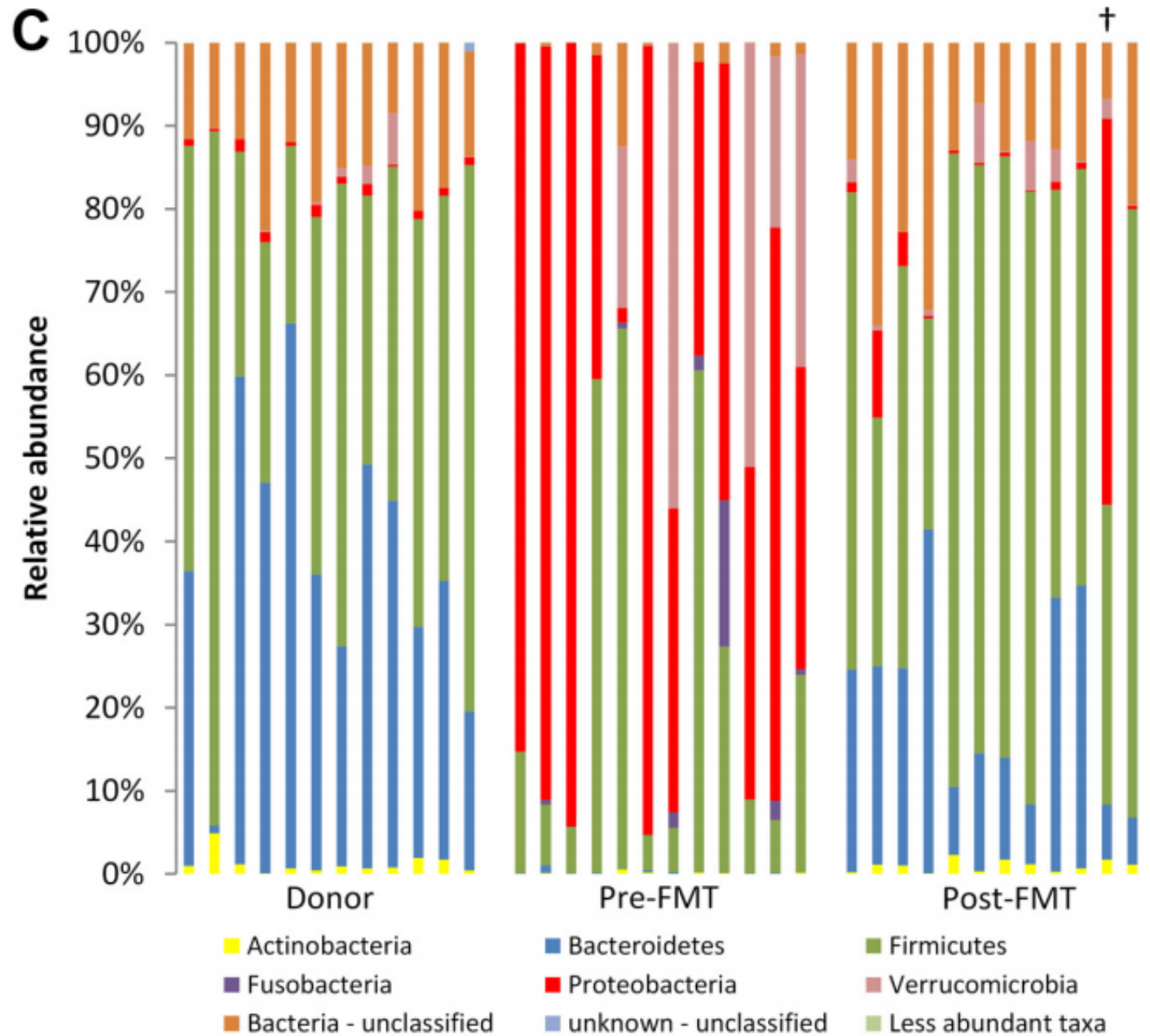
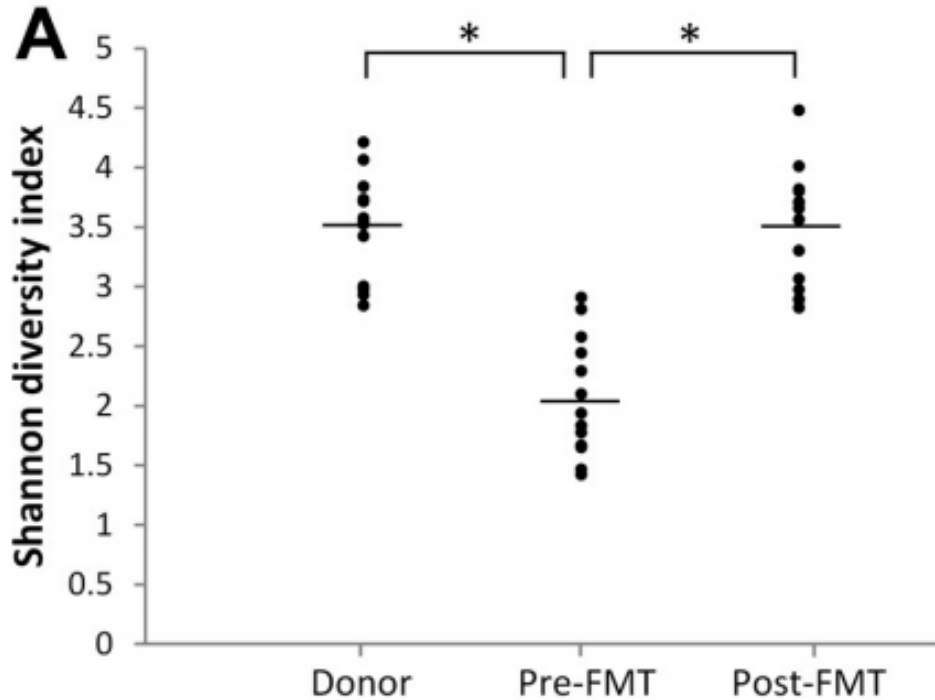
- FMT group: 18/20 (90%) of patients
- Vancomycin group: 5/19 (26%) of patients

Mechanisms of FMT in treating RCDI

- Characteristics of rCDI patient's gut microbiota:
 - Loss of microbial diversity
 - ↓ Abundances of Bacteroidetes and Firmicutes
 - ↑ Abundances of Proteobacteria
(Weingarden et al., 2015)
- FMT normalizes microbial diversity & community structure of patient's gut microbiota
 - Normalization was observed as early as 24 h after FMT
(Weingarden et al., 2014)

Study by Weingarden et al., 2015

- 12 patients & 2 donors
- 16S rRNA sequence characterization on stool sample



Mechanisms of FMT in treating rCDI

Two mechanistic categories

Direct interaction of donor gut microbiota with *C. difficile*

- Antimicrobial peptides
- Bile-acid-mediated inhibition

Microbiota mediated effects on host immune responses

(Khoruts and Sadowsky, 2016)

Mechanisms of FMT in treating rCDI

- Antimicrobial peptides — Bacteriocins

Thuricin CD
(Rea et al., 2010)

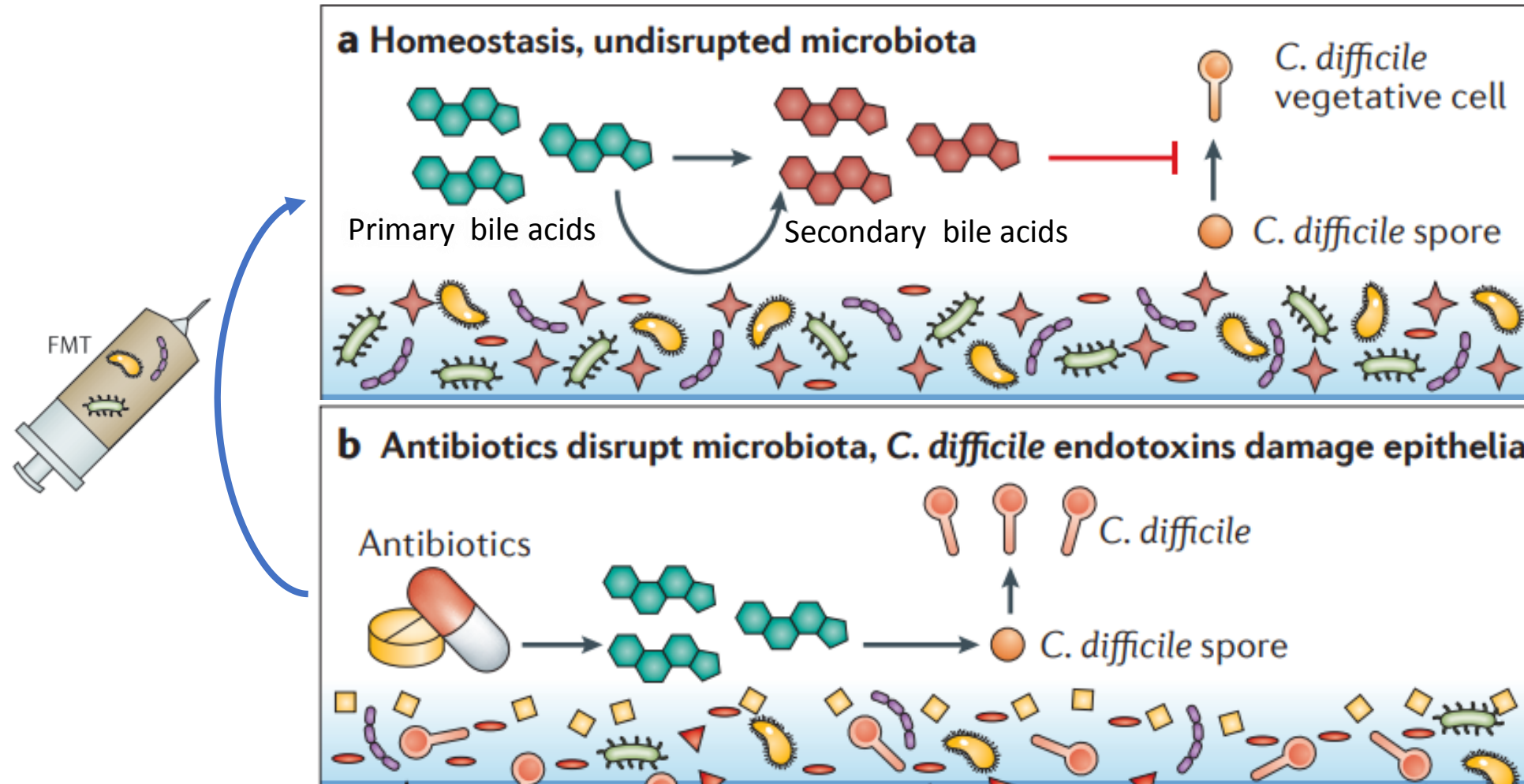
- Produced by *Bacillus thuringiensis*
- Narrow spectrum and highly effective against *C. difficile*

Nisin
(Lay et al., 2016)

- Produced by *Lactococcus lactis*
- Inhibits *C. difficile* vegetative cells growth and spore germination

Mechanisms of FMT in treating rCDI

- Bile-acid-mediated inhibition



(Modified from Figure 1. Khoruts & Sadowsky, 2016)

Challenges faced by FMT

Safety

Applications

Replacement

Challenges faced by FMT—Safety

- Short term adverse effects
 - Minor events
 - e.g. Diarrhoea, abdominal cramps, belching, and constipation
 - Common; Resolve within hours post-FMT
 - Serious events
 - Rare
 - Procedure-related
 - Endoscopy: Perforation and bleeding
 - Sedation: Aspiration
 - Infectious complications
 - Uncertain relationship to FMT

(Cammara et al., 2015;
Khoruts and Sadowsky, 2016;
Weingarden et al., 2013)

Challenges faced by FMT—Safety

- Potential Long-term effects
 1. Possible transmission of infectious agents through FMT
 - e.g. Hepatitis C and HIV
 - Not examined in short-term follow-up
 - Other possible unrecognized infectious agents
 2. Induction of chronic disease related to alterations of microbiota
 - Disease/condition related to gut microbiota:
Obesity, diabetes, colon cancer etc.

To assess the long-term safety of FMT,

Clinical follow-up of patients over years

Analysis of banked donor and recipient specimens

} are required

(Cammara et al., 2017; Kelly et al., 2015)

Challenges faced—Applications other than rCDI

- Refractory CDI
 - Pooled results of 7 case-series studies (Drekonja et al., 2015)
Overall resolution rate 55%, range 0-100%
- Inflammatory bowel disease (IBD)
 - In a systematic review included 53 studies (Paramsothy et al., 2017)
36% [201/555] of ulcerative colitis
50.5% [42/83] of Crohn's disease
patients achieved clinical remission

Challenges faced—Applications other than rCDI

- Obesity
 - Randomized control study of Vrieze et al., 2012
 - Obese male subjects received FMT of allogenic (lean donor) or autologous (self) stool material
 - After 6 weeks, the allogenic group had
 - Improved insulin sensitivity
 - Increased diversity in gut microbiota
 - Increased abundance of butyrate-producing bacteria

Challenges faced—Applications other than rCDI

- Promising preliminary data
- Not efficacious as treating rCDI
- Large randomized controlled studies are required
- Optimization of FMT protocol is required
 - e.g. Antibiotic pretreatment & Multiple rounds of FMT

Challenges faced by FMT—Replacements

- Products with defined microbial consortia are more favoured
- Examples of treating rCDI
 - SER-109
 - Orally administered capsule developed by Seres Therapeutics
 - Contains mixture of bacterial spores purified from human faeces
 - Current in phase 3 trial
 - RePOOPulate (Petrof et al., 2013)
 - Collection of 33 strains isolated from healthy human faeces
 - Administration by infusion through colonoscopy
 - Achieved clinical remission of 6 months in 2 patients

Take-home messages

Fecal microbiota transplantation (FMT) is

- Grafting donor's gut microbiota to a recipient through fecal transplant
- To normalize gut microbiota diversity and composition

FMT has shown to

- Be an highly effective treatment for recurrent CDI
- Have promising results in treating other diseases

FMT lacks

- Assessments on long-term adverse effects
- Optimal protocols

FMT may be replaced by products with defined microbial consortia



Q&A

References

- Cammarota, G., Masucci, L., Ianiro, G., Bibbò, S., Dinoi, G., Costamagna, G., Sanguinetti, M., and Gasbarrini, A. (2015). Randomised clinical trial: faecal microbiota transplantation by colonoscopy vs. vancomycin for the treatment of recurrent *Clostridium difficile* infection. *Aliment. Pharmacol. Ther.* *41*, 835–843.
- Cammarota, G., Ianiro, G., Tilg, H., Rajilić-Stojanović, M., Kump, P., Satokari, R., Sokol, H., Arkkila, P., Pintus, C., Hart, A., et al. (2017). European consensus conference on faecal microbiota transplantation in clinical practice. *Gut* *66*, 569–580.
- Cohen, S.H., Gerding, D.N., Johnson, S., Kelly, C.P., Loo, V.G., McDonald, L.C., Pepin, J., Wilcox, M.H., Society for Healthcare Epidemiology of America, and Infectious Diseases Society of America (2010). Clinical practice guidelines for *Clostridium difficile* infection in adults: 2010 update by the society for healthcare epidemiology of America (SHEA) and the infectious diseases society of America (IDSA). *Infect. Control Hosp. Epidemiol.* *31*, 431–455.
- Drekonja, D., Reich, J., Gezahegn, S., Greer, N., Shaukat, A., MacDonald, R., Rutks, I., and Wilt, T.J. (2015). Fecal Microbiota Transplantation for *Clostridium difficile* Infection: A Systematic Review. *Ann. Intern. Med.* *162*, 630–638.
- Eiseman, B., Silen, W., Bascom, G.S., and Kauvar, A.J. (1958). Fecal enema as an adjunct in the treatment of pseudomembranous enterocolitis. *Surgery* *44*, 854–859.
- Kelly, C.R., Kahn, S., Kashyap, P., Laine, L., Rubin, D., Atreja, A., Moore, T., and Wu, G. (2015). Update on Fecal Microbiota Transplantation 2015: Indications, Methodologies, Mechanisms, and Outlook. *Gastroenterology* *149*, 223–237.
- Khoruts, A., and Sadowsky, M.J. (2016). Understanding the mechanisms of faecal microbiota transplantation. *Nat. Rev. Gastroenterol. Hepatol.* *13*, nrgastro.2016.98.
- Lay, C.L., Dridi, L., Bergeron, M.G., Ouellette, M., and Fliss, I. (2016). Nisin is an effective inhibitor of *Clostridium difficile* vegetative cells and spore germination. *J. Med. Microbiol.* *65*, 169–175.

References

- Li, Y.-T., Cai, H.-F., Wang, Z.-H., Xu, J., and Fang, J.-Y. (2016). Systematic review with meta-analysis: long-term outcomes of faecal microbiota transplantation for *Clostridium difficile* infection. *Aliment. Pharmacol. Ther.* 43, 445–457.
- van Nood, E., Vrieze, A., Nieuwdorp, M., Fuentes, S., Zoetendal, E.G., de Vos, W.M., Visser, C.E., Kuijper, E.J., Bartelsman, J.F.W.M., Tijssen, J.G.P., et al. (2013). Duodenal Infusion of Donor Feces for Recurrent *Clostridium difficile*. *N. Engl. J. Med.* 368, 407–415.
- Paramsothy, S., Paramsothy, R., Rubin, D.T., Kamm, M.A., Kaakoush, N.O., Mitchell, H.M., and Castaño-Rodríguez, N. (2017). Faecal Microbiota Transplantation for Inflammatory Bowel Disease: A Systematic Review and Meta-analysis. *J. Crohns Colitis* 11, 1180–1199.
- Petrof, E.O., Gloor, G.B., Vanner, S.J., Weese, S.J., Carter, D., Daigneault, M.C., Brown, E.M., Schroeter, K., and Allen-Vercoe, E. (2013). Stool substitute transplant therapy for the eradication of *Clostridium difficile* infection: ‘RePOOPulating’ the gut. *Microbiome* 1, 3.
- Rea, M.C., Sit, C.S., Clayton, E., O’Connor, P.M., Whittal, R.M., Zheng, J., Vederas, J.C., Ross, R.P., and Hill, C. (2010). Thuricin CD, a posttranslationally modified bacteriocin with a narrow spectrum of activity against *Clostridium difficile*. *Proc. Natl. Acad. Sci. U. S. A.* 107, 9352–9357.
- Schwan, A., Sjölin, S., Trottestam, U., and Aronsson, B. (1983). Relapsing *clostridium difficile* enterocolitis cured by rectal infusion of homologous faeces. *Lancet Lond. Engl.* 2, 845.
- Vrieze, A., Van Nood, E., Holleman, F., Salojärvi, J., Kootte, R.S., Bartelsman, J.F.W.M., Dallinga–Thie, G.M., Ackermans, M.T., Serlie, M.J., Oozeer, R., et al. (2012). Transfer of Intestinal Microbiota From Lean Donors Increases Insulin Sensitivity in Individuals With Metabolic Syndrome. *Gastroenterology* 143, 913–916.e7.

References

- Weingarden, A., González, A., Vázquez-Baeza, Y., Weiss, S., Humphry, G., Berg-Lyons, D., Knights, D., Unno, T., Bobr, A., Kang, J., et al. (2015). Dynamic changes in short- and long-term bacterial composition following fecal microbiota transplantation for recurrent *Clostridium difficile* infection. *Microbiome* 3, 10.
- Weingarden, A.R., Hamilton, M.J., Sadowsky, M.J., and Khoruts, A. (2013). Resolution of Severe *Clostridium difficile* Infection Following Sequential Fecal Microbiota Transplantation. *J. Clin. Gastroenterol.* 47, 735–737.
- Weingarden, A.R., Chen, C., Bobr, A., Yao, D., Lu, Y., Nelson, V.M., Sadowsky, M.J., and Khoruts, A. (2014). Microbiota transplantation restores normal fecal bile acid composition in recurrent *Clostridium difficile* infection. *Am. J. Physiol. - Gastrointest. Liver Physiol.* 306, G310–G319.